Cellular, molecular, and clinical mechanisms of action of deep brain stimulation—a systematic review on established indications and outlook on future developments

Martin Jakobs1,2,*, Anton Fomenko2,*, Andres M Lozano2 & Karl L Kiening1

Abstract

Deep brain stimulation (DBS) has been successfully used to treat movement disorders, such as Parkinson’s disease, for more than 25 years and heralded the advent of electrical neuromodulation to treat diseases with dysregulated neuronal circuits. DBS is now superseding ablative techniques, such as stereotactic radiofrequency lesions. While serendipity has played a role in developing DBS as a therapy, research during the past two decades has shown that electrical neuromodulation is far more than a functional lesion that can be switched on and off. This understanding broadens the field to enable new types of stimulation, clinical indications, and research. This review highlights the complex effects of DBS from the single cell to the neuronal network. Specifically, we examine the electrical, cellular, molecular, and neurochemical mechanisms of DBS as applied to Parkinson’s disease and other emerging applications.

Keywords  ciruuptopathies; deep brain stimulation; movement disorders; neuromodulation; Parkinson’s disease

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See the Glossary for abbreviations used in this article.

Introduction

Deep brain stimulation (DBS) has been an established neurosurgical procedure since the 1990s, and more than 160,000 patients worldwide have been operated (Fig 1). During stereotactic surgery, electrodes are implanted in the subcortical nuclei or fiber bundles of the brain, in order to apply chronic low electric currents to therapeutically alter neural function. DBS is an established treatment for movement disorders such as Parkinson’s disease (PD; Brown et al, 1999; Schuepbach et al, 2013), several forms of tremor (Hubble et al, 1996), and dystonia (Kumar et al, 1999). Other clinical indications are treatment-resistant epilepsy (Fisher et al, 2010) and obsessive–compulsive disorder (OCD; Greenberg et al, 2006). Before the advent of DBS, movement disorder surgery encompassed ablative techniques using mostly radiofrequency lesions. Stereotactic lesions in the thalamus (“thalamotomy”; Hassler, 1972) and the pallidum (“pallidotomy”; Spiegel & Wycis, 1952) were commonly used, especially before the introduction of levodopa medication to treat PD. To predict the effects of lesioning prior to a thalamotomy in the ventral intermediate (Vim) nucleus of the thalamus, surgeons used high-frequency stimulation, which yielded a sustained and immediately reversible effect on tremor (Benabid et al, 1996). Thus, the idea to alter the function of neuronal tissue with electricity rather than to destroy it was born. This discovery led to the development of fully implantable DBS systems with dual electrodes connected to an implantable pulse generator (IPG) similar to a cardiac pacemaker to deliver long-term and continuous therapy (Fig 2).

DBS is nowadays part of a class of treatments known as neuromodulation, which modifies neural function through application of electrical currents. The effects are for the most part immediate, reversible, adaptable, and titratable without permanently damaging neural tissue. Additionally, contrary to lesioning techniques, DBS can be applied bilaterally without causing severe side effects in movement disorder surgery.

The observation that high-frequency electric stimulation caused the same effects as ablative techniques led to the assumption that DBS inhibits neuronal activity of the stimulated nucleus, causing a functional lesion (Johnson & Vitek, 2011) via depolarization blocks (Meissner et al, 2005; McIntyre & Anderson, 2016). While functional inactivation of neurons and depolarization blocks may contribute to the overall effect, further research revealed that the effects of DBS on the cellular, electrical, molecular, and network level by far surpass the effects of lesions. The current hypothesis is that
**Glossary**

**Action potential (AP)**  
A.k.a. nerve impulse or spike: a propagation of electrical depolarization along the length of a neuron's axon, mainly modulated by voltage-gated on channels. Otherwise known as a nerve impulse.

**Afferent**  
Nerve fibers (axons) that arrive at a certain brain region and convey information from other brain regions via synapses on the dendrites of that target neuron.

**Anhedonia**  
A loss of interest or reduced ability to experience pleasure. A core component of several depression-related disorders.

**Antidromic**  
Propagation of an action potential along an axon in the opposite direction of usual (orthodromic) conduction. Antidromic excitation of a nerve refers to travel of an electrical signal away from axon terminals and toward the cell body.

**Bradykinesia**  
A.k.a. slowing of movement: Along with tremor and rigidity, bradykinesia is a cardinal symptom of certain neurodegenerative disorders such as Parkinson's disease. Bradykinesia can sometimes also be a side effect of certain medications.

**Cyclic voltammetry**  
An electrochemical analytical technique that allows monitoring of neurotransmitter concentrations by probing different brain regions with an electrode. Neurotransmitters detected using this technique include dopamine, serotonin, and norepinephrine.

**Deafferentation**  
A condition resulting from the elimination or interruption of nerve fibers (afferents) that carry sensory information to the brain. Potential causes of this condition are disease, injury, or in an experimental context, an intentional lesion caused to study resulting effects on the peripheral and central nervous system.

**Deep brain stimulation (DBS)**  
A surgical procedure involving implantation of an implantable pulse generator (IPG), leads, and electrode(s) to deliver chronic electrical stimulation of brain region(s). DBS is used either for therapy for disorders affecting the nervous system, especially movement disorders, and has been approved in most countries for the treatment of Parkinson's disease, essential tremor, and dystonia. Experimental applications include other neurodegenerative conditions such as Alzheimer's disease, as well as psychiatric disorders such as depression.

**Depolarization block**  
An electrical event within a neural membrane caused by sustained input currents, especially at strong intensities. For example, during seizures, sustained firing of neurons may lead to a depolarization block mediated by persistent sodium currents, which limits the maximum firing rate of the cells.

**Dyskinesia**  
An abnormal, involuntary, repetitive movement of a limb or part of the body, for example, a twitch. In context of Parkinson's disease, dyskinesia is a common side effect of long-term use of antiparkinsonian dopaminergic medications, such as levodopa.

**Dystonia**  
A movement disorder in which sustained, involuntary muscle contractions cause painful and abnormally contorted body or limb positions. Dystonia can be either congenital or acquired and is one of the common indications for DBS treatment, with a common target being the globus pallidus interna.

**Effenter**  
Nerve fibers (axons) that exit from a certain brain region and convey information from the target neurons via synapses on the dendrites of other brain regions.

**Entorhinal cortex (EC)**  
A cortical brain region located inferior to the rhinal sulcus within the medial temporal lobe. Purported to serve as a hub for memory-related pathways and a site of experimental DBS for Alzheimer's disease, especially in animal models.

**Implantable pulse generator (IPG)**  
A small pacemaker-like implantable medical device that both controls and provides electrical power necessary for a DBS system. Typically implanted under the chest skin, the IPG contains either a rechargeable or single-use battery along with one or more output cables that connect to the stimulating electrodes.

**Magnetoencephalography (MEG)**  
A non-invasive functional brain imaging technique that detects the minuscule electromagnetic fluctuations emitted by the human brain, allowing for monitoring of awake brain activity in multiple cortical brain regions.

**Orthodromic**  
The typical direction of an action potential's propagation, namely away from the neuron cell body, or soma, and toward the axon terminals. Its opposite is antidromic.

**Palidotomy**  
A surgical procedure involving targeted destruction of part of the globus pallidus internus. Sometimes used as an alternative to DBS, this procedure is more commonly used to treat severe cases of levodopa-induced dyskinesia, a common side effect of antiparkinsonian medications.

**Positron emission tomography (PET)**  
A functional brain imaging technique that uses intravenously injected tracers that emit positrons (gamma radiation) to assess metabolic activity. Different tracers are available for different organs or conditions. Gamma radiation is detected over time (dynamic) or at a given point in time after injection of the tracer (static). Results are usually visualized with a three-dimensional CT scan.

**Stereotactic surgery**  
Stereotactic surgery is a minimally invasive form of neurosurgery that utilizes a virtual three-dimensional coordinate system to localize small and often deep-seated brain region so be treated. A stereotactic frame may be placed on the patient head which is utilized as a base for imaging and to anchor the surgical device to bring electrodes, probes, or catheters to this target area. So-called frameless systems utilize head-mounted devices to reach the deep-seated target regions.

**Stimulating electrode**  
A thin metallic conductor of electricity that is implanted within the brain at the desired stimulation target. Typically insulated along most of their length, DBS electrodes contain small uninsulated segments near the stimulating tip. Some DBS systems allow for selection of bipolar or unipolar electrode channel configurations depending on which segments are chosen to conduct electrical current.

**Striatum**  
A collection of nuclei within the basal ganglia comprising the caudate, putamen, and nucleus accumbens. A striped appearance on brain imaging explains this region's etymology from the Latin "striped".

**Subcortical**  
Located within the white matter tracts below the cerebral cortex.

**Subthalamic nucleus (STN)**  
The STN is a small paired lens-shaped nucleus situated inferior to the thalamus. An important coordinator of the output flow from the basal ganglia, the STN has been found to contain "pacemaker neurons" that have spontaneous oscillating and synchronous activity, potentially related to the resting tremor found in Parkinson's patients. Accordingly, the STN is a common site for therapeutic DBS for this disease. The STN is composed of a motor region which is targeted in DBS surgery, but also contains parts connected to the limbic system and to the associative system. Both regions are thought to be avoided during DBS surgery as stimulation of these areas evokes side effects like psychiatric symptoms (i.e., depression/mania for the limbic part) or double vision (for the associative part).

**Thalamotomy**  
A surgical procedure involving targeted destruction of part of the thalamus, a key brain region that relays sensorimotor information to the cerebral cortex. Introduced in the 1950s, thalamotomies are used to alleviate the tremor symptoms of several movement disorders such as Parkinson's disease and essential tremor. Nowadays, this procedure can be performed via traditional surgery or noninvasively using high-intensity focused ultrasound thalamotomy.

**Voltammetry**  
An electroanalytical method to identify the substrates of sample by analyzing its unique conductive properties.
pathologically altered and dysfunctional neuronal circuits, or circuitopathies (Lozano & Lipsman, 2013), are treatable with DBS and that electrical stimulation can alter these neuronal circuits back to a more physiological state (Dostrovsky & Lozano, 2002; Gubellini et al., 2009; Johnson & Vitek, 2011; McIntyre & Anderson, 2016).

In this review, we will examine the immediate, short-term, and long-term effects of DBS on a cellular, network, and molecular level and highlight investigational and new indications for DBS. A comprehensive literature search was performed and included original articles on in vitro, in vivo, and clinical research and selected review articles.

Electrical effects at the stimulation target

The electrical excitability of neural tissue around the electrode and the effects of stimulation are determined by several factors: (i) the proportion of gray and white matter structures (i.e., cell bodies or axons; Nowak & Bullier, 1998; Ranck, 1975; Dostrovsky & Lozano, 2002; Udupa & Chen, 2015); (ii) the type of ion channels on the cell membrane of a soma or axon (Hoang et al., 2017); (iii) the diameter and degree of myelination of axons (Ranck, 1975; Nowak & Bullier, 1998; Gubellini et al., 2009; Carron et al., 2013); (iv) the orientation of axons in relation to the electrode (Ranck, 1975; Gubellini et al., 2009; Hoang et al., 2017); and (v) the proximity of the electrode to other structures that can affect the electrical field (Nowak & Bullier, 1998; Dostrovsky & Lozano, 2002; Carron et al., 2013).
Although the electrical effects of clinically applied DBS cannot be
generalized, several principal cellular effects have been consistently
observed (Fig 3). Since stimulation of the subthalamic nucleus
(STN) for PD has been the best studied application in experiments,
the local and network effects of stimulation will be presented in the
case of treating PD (see Fig 4). Stimulation at other targets for
other indications may underlie different principles or varying contri-
butions of these principles.

Standard clinically effective STN-DBS settings comprise the tonic
continuous application of monophasic cathodic impulses at a
frequency of 130 Hz and a pulse width of 60 μs at an amplitude of
0.8–2.0 mA. Chronaxie and rheobase are two means to describe
electrical excitability of neurons and their different components.
Rheobase is the amount of current (of infinite duration) necessary
to depolarize a neuron and cause an action potential (AP; Nowak &
Bullier, 1998; Dostrovsky & Lozano, 2002). Compared to cell bodies,
axons have a lower rheobase and therefore show higher levels of
excitability (Nowak & Bullier, 1998). Chronaxie is the amount of
time needed for a stimulus double the strength of the rheobase to
evoke an AP in a neuron (Holsheimer et al, 2000). Chronaxies for
neuronal cell bodies and dendrites are much longer (1–10 ms)
compared to axons (Ranck, 1975), especially myelinated axons with
a larger diameter, which have chronaxies between 30 and 300 μs
(Holsheimer et al, 2000).

Since a single DBS pulse can influence neuronal activities for
several milliseconds—7–8 ms in some cases—frequencies with a
lesser inter-stimulus interval (> 125 Hz) will prevent a neuron from
fully returning to its base activity (Baker et al, 2002; Montgomery &
Gale, 2008). It is largely believed that clinically effective high-
frequency DBS preferentially acts on large myelinated axons, depo-
larizes them, and causes APs (Montgomery & Gale, 2008; Johnson &
Vitek, 2011) roughly at the frequency of the stimulation by opening
voltage-gated sodium channels (McIntyre & Anderson, 2016). APs
have been shown to travel along a stimulated axon in orthodromic
fashion toward the synapse as well as in antidromic fashion toward
the soma (Li et al, 2007; Agnesi et al, 2013; Carron et al, 2013).

Orthodromic APs can then trigger neurotransmitter release at the
synapse and postsynaptic potentials in the efferent target neurons
(Shen et al, 2003). Thus, most of the efferent STN axons fire APs in
accordance with the frequency of the DBS (Johnson & Vitek, 2011;
Humphries & Gurney, 2012) once the stimulation frequency is
roughly twice as high as their intrinsic firing frequency (McIntyre &
Anderson, 2016). However, the generation and propagation of APs
can vary during long-term stimulation (McIntyre & Anderson,
2016). As the volume of tissue activated (VTA) by clinical DBS does
not only contain efferent STN axons but also afferent fibers project-
ing to STN dendrites and cell bodies, inhibitory (GABAergic) and
excitatory (glutamatergic) afferent fibers can be activated and act on
the STN cell body itself (Dostrovsky et al, 2000; Anderson et al,

Cell bodies activated by afferent fibers can generate orthodromic
APs that have been shown to collide with antidromic APs generated
by axonal stimulation. This phenomenon partially cancels out
the intrinsic firing pattern of STN neurons (Magarios-Ascone et al,
2002; Meissner et al, 2005). When antidromic APs reach the soma,
they initially activate it before constant AP influx overrides the
refractory period and prevents the neuron from firing in its previous
pattern (Johnson & Vitek, 2011). Two types of reactions have been

Figure 2. DBS system used for clinical applications.
Clinically used DBS system: The brain electrode delivers the electrical current for
therapeutic purposes. The extension lead connects the brain electrode to the
neurostimulator (internal pulse generator, IPG), the implanted power source.
observed in the soma of STN neurons during stimulation: Type I responses incorporate short phases of depolarization before returning to baseline. Type II responses show a prolonged period of depolarization with or without firing spikes of APs (Shen et al., 2003; Anderson et al., 2004). This constant trend toward depolarization of cell bodies was thought to be a stable depolarization block comparable to the effects of lesioning techniques (Benazzouz & Hallett, 2000; Beurrier et al., 2001; Kringelbach et al., 2007b; Hamani et al., 2017) to silence the targeted cells.

However, it has been shown that depolarization blocks per se are unsustainable states. Cells regain the ability to repolarize over time and fire intrinsic spikes of APs (Ammari et al., 2011). Intermittent firing was observed in tonic or burst patterns before the cell enters a phase of depolarization block again. The intrinsic APs have lower amplitudes, but can still evoke neurotransmitter release at the synapse (Florence et al., 2009; Ammari et al., 2011). Even constantly depolarized membranes in a silenced cell are capable of releasing smaller amounts of neurotransmitters. Depolarization blocks lead to inactivation of voltage-gated sodium ion channels and accumulation of extracellular potassium (Shin et al., 2007). This helps to maintain the resting membrane potential, but has the potential to disturb the conventional mechanisms of repolarization which favors lasting neuronal activity (Florence et al., 2009). Over time, Na⁺/K⁺ ATPases ultimately repolarize the cell and enable neuronal firing again. Overall, the firing rate of STN cell bodies is reduced with high-frequency DBS (Degos, 2005). In summary, DBS therapy seems to inhibit cell bodies and reduce their firing rate while it excites axons and increases their AP frequency.

Astrocytes are another important player in the ion and neurotransmitter microenvironment in the VTA. These glial cells release
neurotransmitters and neuromodulators that influence neurons, which might contribute to the therapeutic long-term effect of DBS (Fenoy et al., 2014). Electrical stimulation can activate astrocytes via calcium ion influx, and activated astrocytes have the capability to release glutamate and adenosine and increase the probability of neurotransmitter release from excitatory or inhibitory presynaptic terminals (Sasaki et al., 2011). This was shown in in vitro experiments: When neuronal synaptic activity was blocked in the thalamus, vesicular glutamate release was still found via astrocytes (Tawfik et al., 2010). Furthermore, astrocytes will shift potassium into the extracellular space and therefore contribute to a microenvironment that supports depolarization block-like states (Sasaki et al., 2011; Barat et al., 2012; Sauleau et al., 2016). The change in the microenvironment of non-neural cells might lead to prolonged plasticity-associated effects even when stimulation is turned off. Although there is some in vitro evidence that astrocytes may be actively involved in the effects and mechanisms of DBS, no clinical in vivo studies have so far been able to prove this theory.

In summary, DBS seems to uncouple STN neurons from its axons and cause a functional deafferentation from both efferent and afferent structures (Dejean et al., 2009; Moran et al., 2011). Different factors may contribute to the spike activity of inhibited STN neurons: intermittent repolarization during depolarization blocks or preponderance of activation of excitatory afferent projections to the STN neurons. The efferent synapse, however, will not at all times be entrained by the frequency of the stimulation or be completely silenced. Considering the temporospatial aspects of DBS, not all target cells in the VTA may act in complete synchronicity. Some cells may be silenced, and others will be entrained in tonic firing or fire in intermittent bursts. Overall, the axons of the stimulated target are “hijacked” by high-frequency stimulation, and a new firing pattern is encoded (Benazzouz & Hallett, 2000; Dostrovsky & Lozano, 2002) into the circuit that overwrites the pathological firing pattern in circuitopathies. The collective intrinsic spiking generated by the soma in tonic or burst fashion will cause irregular synaptic and injection of noise in the STN efferents and target neurons (Anderson et al., 2004; Moss et al., 2004; Hamani et al., 2012).

**Electrical effects in the neuronal network**

Owing to practical and ethical considerations, human studies on the electrical effects of DBS are most often limited to a single target as the standard of clinical application. These studies either encompass intraoperative measurements via microelectrode recording and microstimulation or recordings from temporarily externalized electrodes after implantation. Multi-target studies are usually based on in vivo animal experiments or in vitro brain slices. Most of the knowledge about network effects therefore comes from experimental animal studies.

The basal ganglia network in healthy individuals entails several neural circuits and both excitatory and inhibitory connections, the correct balance of which enables the appropriate and correct amount of movement (Fig 5A). As a circuitopathy, PD is characterized by the loss of dopaminergic neurons in the substantia nigra pars compacta (SNc) associated with misfolded α-synuclein proteins and the formation of Lewy bodies (Kalia et al., 2013). This dopaminergic loss leads to various and widespread changes in the basal ganglia circuits and cause the main motor symptoms of PD (Fig 5B): bradykinesia, rigidity, and resting tremor. In this pathologically disrupted neural circuit, the electric and functional uncoupling of stimulated STN neurons inhibits the influence of afferent nerves (globus pallidus externus (GPe; Fig 5C), SNc, and the motor cortex through the hyperdirect pathway (Nambu et al., 1996). Uncoupling the STN from the SNc makes patients with STN-DBS less dependent on endogenous and exogenous dopamine.

The main output structures for glutamatergic STN neurons are the globus pallidus pars internus (GPI) and the substantia nigra pars reticulata (SNr; Fig 5A). High-frequency DBS has been shown to alter neuronal activity in all parts of the basal ganglia, including its main release station within the thalamus (Dorval et al., 2008). In the GPI, STN-DBS causes significant excitation and inhibition in different neurons, while the firing frequency is strongly entrained by the DBS frequency (Hahn & McIntyre, 2010). The GPI showed less activity under STN-DBS (Hashimoto et al., 2003) with increased thalamocortical activity (Fig 5C). STN-DBS appears to overwrite one form of pathologic activity with an artificially created “noisy” activity, which is less detrimental to the motor circuit.

The pathological basal ganglia network activity in PD is represented by increased synchrony and rhythmicity of neural activity and oscillations. Neural oscillations can be subdivided into different frequency bands: δ (delta: 1–4 Hz), θ (theta: 4–7 Hz), α (alpha: 7–13 Hz), β (beta: 14–30 Hz), and χ (gamma: 30–100 Hz; Başar et al., 2013). Theta and especially beta oscillations are thought to be antikinetin (Brown & Williams, 2005) and physiologically maintain the status quo of the resting network (Johnson & Vitek, 2011). Gamma oscillations seem to be associated with hyperkinetic states before and during movements, but also with dyskinetic hypermobility in PD.

**Figure 4. Intraoperative DBS electrode targeting.**

STN-DBS: DBS electrode implanted in the left dorsolateral subthalamic nucleus (green) in relation to the substantia nigra (blue) and the red nucleus (red).
Overall, PD patients show an increase in beta and a reduction in gamma oscillations (Silberstein et al., 2003; Kühn et al., 2006) in most basal ganglia structures, such as the striatum, STN, pallidum, and the motor cortex, which correlates with states of bradykinesia and rigidity (Basar et al., 2013; Little & Brown, 2014). Further synchrony has been detected as the phase of a lower-frequency (beta) oscillation is able to control the amplitude of a higher-frequency (gamma) oscillation (Moran et al., 2011; De Hemptinne et al., 2015). This phase–amplitude coupling (De Hemptinne et al., 2015) has been observed in the motor cortex of PD patients with bradykinesia.

Thus, the current hypothesis of basal ganglia networks in PD is that the overall output to the thalamus and motor cortex is decreased and the circuit is caught in a hyper-synchronized oscillatory state of entropy (Dorval et al., 2008; Anderson et al., 2015; Hoang et al., 2017). The efferent motor synapses of the STN during DBS are hypothesized to be depleted of their neurotransmitters and thereby filter low-frequency oscillations (McIntyre & Anderson, 2016). Low-frequency DBS (20 Hz) appears to increase the amount of synchronization, whereas high-frequency DBS (> 70 Hz) suppresses such activity in the GPi (Brown et al., 2004), similar to the administration of levodopa (Dorval et al., 2008; Hoang et al., 2017).

The injection of postsynaptic noise and the decoupling of the STN neurons has been shown to reduce the power of beta oscillations, reduce phase/amplitude coupling, and decrease the entropy in the firing pattern of cortical neurons (Little & Brown, 2014; De Hemptinne et al., 2015; McIntyre & Anderson, 2016). This new firing pattern likely does not recreate the physiological firing pattern, but replaces the more deleterious intrinsic pattern of the corresponding circuitopathy. The overall network effect of STN-DBS is an increased activity in the basal ganglia output from the thalamus to the motor cortex along with reduced synchrony in lower-frequency oscillations.

**Neurochemical effects**

DBS also seems to change the neurochemical milieu of neurons and neuronal tissue (Gondard et al., 2015; Udupa & Chen, 2015; Fischer
et al., 2017; Torres-Sanchez et al., 2017). Although the immediate effects of DBS are an altered firing pattern of neural circuits, these are often followed by changes in neurotransmitter dynamics and protein expression, which inevitably affect the behavior of the network in the longer term (McIntyre & Hahn, 2010; Fig 6).

**Dopamine**

Dopamine is a well-known therapeutic for treating movement disorders, such as PD, that are characterized by reduced dopamine production (Perlmutter & Mink, 2006). Animal models were thus used to quantify dopaminergic flux during and after DBS. In both anesthetized (Bruet et al., 2001; Meissner et al., 2001) and awake rats, STN-DBS increases extracellular striatal dopamine in wild-type and dopamine-depleted animals (Meissner et al., 2002). Concomitant circling behavior was demonstrated during DBS, which suggests activation of gammaaminobutyric acid (GABA) receptors in the STN (Murur & Pazo, 1993). Complementary electrophysiologic studies in STN-stimulated wild-type rodents showed time-locked increases in striatal dopamine efflux, as well as up to an 11-fold chronic increase in dopamine after cessation of stimulation at 50 Hz and 200 μA (Lee et al., 2006).

In contrast to these animal studies, the majority of studies in PD patients undergoing STN-DBS failed to show increased dopamine release, in spite of tremor amelioration during stimulation (Hilker et al., 2003; Strafella et al., 2003; Thobois et al., 2003). An isolated study in three PD patients, however, did suggest increased dopamine concentrations after stimulation (Nimura et al., 2005). The discrepancy in these experiments may result from differences in the precise subregion of the targeted STN (motor, affective, limbic part), which may affect the magnitude of dopamine release. Accordingly, in rhesus macaques, cyclic voltammetry demonstrated STN-DBS-evoked dopamine release in the caudate and putamen, with maximal concentration effects seen during stimulation of the posterior and lateral regions of the STN (Gale et al., 2013; Min et al., 2016). Notably, dopaminergic fibers are unmyelinated, which lowers their susceptibility to electrical stimulation compared to myelinated sites of DBS and may help explain the discrepancy in results (Kringelbach et al., 2007b). In summary, animal models have suggested DBS-induced dopaminergic facilitation, though the findings have not been robustly substantiated in patients. This may be related to the fact that dopamine is greatly depleted in patients with advanced PD and thus unavailable for release in response to DBS.

**Noradrenaline**

Noradrenaline (NA) might be involved in the mechanisms of action of DBS given the noradrenergic projections to the STN originating from the locus coeruleus. Depletion of NA and its metabolites within the limbic system has been implicated in both movement disorders and psychiatric pathology (Remy et al., 2005; Delaville et al., 2011). Specifically, NA is a key mediator of the cortico-striato-thalamo cortical pathway, components of which are targeted in DBS for obsessive-compulsive disorder (OCD) and major depressive disorder (MDD). Accordingly, bilateral nucleus accumbens (NAcc) DBS in rats yielded a sustained rise in dopamine and NA in the orbitofrontal cortex of up to 166% from baseline (Van Dijk et al., 2012). However, the same group found no increases in monoamine concentration near the stimulation site itself, suggesting that distal NA release dominates (Van Dijk et al., 2011). DBS of the infralimbic prefrontal cortex in a rodent depression model showed increased NA expression at the stimulation site, along with antidepressive effects (Jiménez-Sánchez et al., 2016). More recently, DBS of the vmPFC (ventromedial prefrontal cortex: a rodent homolog to the human SCC) in a depression model helped to illuminate the electrophysiological mechanisms of DBS-induced amine production (Torres-Sanchez et al., 2018). Along with antidepressive effects, stimulation led to increased spontaneous firing of the locus coeruleus and increased expression of tyrosine hydroxylase, which catalyzes the biochemical synthesis of NA and other amines (Torres-Sanchez et al., 2018).

**Serotonin**

The serotonergic system is involved in prefrontal-raphe limbic pathways, and its dysfunction has been implicated in mood disorders such as MDD. Recently, vmPFC-DBS was found to increase serotonin levels by 33–55% in both wild-type and serotonin transporter knockout mice while inducing antidepressive-like behavior (Bregman et al., 2017). Similar experiments in rats showed up to 50% increase in serotonin in the hippocampus following vmPFC-DBS (Boggio et al., 2005), as well as preferentially reversing anhedonia-like behavior in animals with an intact serotonergic system (Hamani et al., 2012).

Recently, renewed appreciation of serotonin’s role in the motor and non-motor symptoms of PD has stimulated research into the serotonergic effects of DBS treatment for movement disorders. In a haloperidol-induced tardive dyskinesia rat model, STN-DBS resulted in reduced serotonin release from the dorsolateral striatum, correlating with reduced dyskinetic movements in the animals (Creed et al., 2012). However, the same experiment found that when serotonin reduction is pharmacologically blocked, STN-DBS was still able to rescue the phenotype, suggesting that several parallel therapeutic
mechanisms are involved (Creed et al., 2012). Other experiments have similarly found serotonin-lowering effects of DBS in the hippocampus, prefrontal cortex (Navailles et al., 2010), and the striatum (Tan et al., 2013). A potential mechanism is electrical inhibition of dorsal raphe neurons that reside in the largest serotonergic nucleus of the brain (Temel et al., 2007). Though the STN provides few direct projections to the dorsal raphe proper, it does innervate the lateral habenula, which provides convergent input to the dorsal raphe nucleus, thereby suggesting a possible indirect serotonin-modulating mechanism for STN-DBS (Kalén et al., 1989; Temel et al., 2009).

Gammaaminobutyric acid
Clinical studies have implicated activation of inhibitory GABAergic pathways as part of the therapeutic effects of electrical brain stimulation (Lozano & Eltahawy, 2004). For instance, intraoperative microstimulation of the GPi in PD patients was found to activate afferent striatal and GPe axons, resulting in GABA release and concurrent GPi inhibition (Dostrovsky et al., 2000). GPi, being the main output structure of the basal ganglia to the thalamus, therefore showed reduced GABAergic activity under STN-DBS (Hashimoto et al., 2003; Miocinovic, 2006). Electrophysiological studies in primates revealed this inhibition to likely be mediated via GABAA and GABAB receptors (Chiken & Nambu, 2013). Similarly, in an STN-DBS cohort of PD patients, both levodopa administration and electrical stimulation caused a decrease in GABA content in the motor thalamus, suggesting similar mechanism of motor symptom alleviation (Stefani et al., 2011). GABA levels were increased in the SNr during STN-DBS, which showed normalized reduced neural activity (Tai, 2003) owing to decreased glutamatergic excitation. Since most DBS targets have both excitatory and inhibitory efferents and afferents, the overall effect on circuit behavior and phenotype is likely complex and is influenced by the relative distribution of these fibers (Lozano & Eltahawy, 2004; Montgomery & Gale, 2008).

Neurochemical effects of electrode implantation
As previously mentioned, electrical effects of DBS underlie not only spatial but also temporal aspects. An animal study showed that implantation of a DBS electrode in the dorsal hippocampus has widespread and early effects on the brain even without electrical stimulation. The cellular effects were most pronounced after 2 weeks with microgliosis and astrocytosis; most of these cells were generated within 3 days after implantation (Song et al., 2013). The cytokine milieu showed expression of the pro-inflammatory cytokine IL-12, significant expression of granulocyte-colony-stimulating factor (G-CSF) along with increased expression of epidermal growth factor (EGF) and brain-derived neurotrophic factor (BDNF; Song et al., 2013). Immunohistochemical staining showed an increased pattern of doublecortin (DCX) positive cells—a marker for immature neurons—indicating neurogenesis in the hippocampal subgranular zone (Song et al., 2013). These early and fluid effects of electrode insertion have clinical implications as many centers do not start stimulation in the early phase after the surgery while the microenvironment around the electrode is still adapting to the effects of implantation.

In a prospective study, all patients showed a certain degree of peri-electrode edema in MRI scans taken 3–20 days after the procedure and a rate of microhemorrhages of more than 30% (Borellini et al., 2018). This can be associated with temporary neurological deficits, states of confusion, or seizures (Deogaonkar et al., 2011; Arrocho-Quinones & Pahapill, 2016). Edema has not only been shown to be present at the tip of the electrode but also along the trajectory (Deogaonkar et al., 2011). Edema usually resolves after 4–6 weeks and, so far, there is no evidence that the presence or the size of edema correlates with the extent or duration of the insertional effect.

Microhemorrhages, edema, and other reactions around the electrode can change the tissue impedance, especially within the first days after surgery, but also over longer terms (Hartmann et al., 2015). When utilizing a voltage-controlled DBS system, impedance changes theoretically cause varying currents and therefore VTAs, which can lead to under-, or more common, overstimulation, and side effects. Current-controlled DBS systems that adapt the voltage to the impedances may therefore be more suitable for earlier programming (Picillo et al., 2016). However, to date no evidence-based guidelines for when to start stimulation exist. Some centers prefer early programming after surgery while others wait 4 weeks or more (depending on the microlesional effect) until they begin stimulation (Kupsch et al., 2011; Picillo et al., 2016).

To determine long-term effects on the tissue surrounding the electrode, cadaver studies of PD patients with STN-DBS showed an up to 1-mm-thick capsule around the electrode with a characteristic three-layer structure. The inner layer was a thin 25-μm membrane of fibrous tissue. The next layer was a 500-μm-thick rim of fibrillary gliosis followed by an up to 1,000-μm-thick layer consisting of reactive astrocytes (Reddy & Lozano, 2017). Signs of a microhemorrhage in the form of hemosiderin as a part of the insertional or microsional effect were also present as well as signs of immune response in the form of t-lymphocytes and multinucleated giant cells (Reddy & Lozano, 2017). Whether this is attributed to direct effects of the stimulation or a foreign-body reaction is unclear.

Vascular changes have been observed too. Endothelial wall thickness was found to be reduced in PD patients compared to non-PD subjects; however, patients with STN-DBS showed increased thickness even compared to non-PD subjects (Pienaar et al., 2015). Although wall thickness increased with STN-DBS, tight junction proteins were found to be reduced compared to non-DBS PD patients (Pienaar et al., 2015). This, together with the finding of an increased capillary density, and increased expression of vascular endothelial growth factor (VEGF) and glial-derived neurotrophic factor (GDNF), led to the hypothesis that STN-DBS changes the microvasculature and causes sprouting of new more stable vessels compared to PD patients without DBS therapy (Pienaar et al., 2015).

Longer-term effects
Changes in neurotransmitter release, expression, and receptor dynamics after DBS can have network and biochemical effects that transcend the time and spatial zone of stimulation (Deniau et al., 2010; McIntyre & Hahn, 2010). This finding has spurred pre- and clinical research into novel applications of DBS, such as treatment of dementia-related disorders (Laxton et al., 2010; Hescham et al., 2013; Sankar et al., 2015), stroke (Elias et al., 2018), pain syndromes
was highly predictive for clinical outcome, corroborating evidence from the supplementary motor area and primary motor cortex, which includes subgenual cingulate and mid-anterior orbitofrontal cortex, associated with pain relief (Kringelbach et al., 2007a). More recently, MEG was used to show that DBS of the anterior cingulate cortex (ACC) for chronic pain induced long-term functional changes in a pain-associated network for up to 1 year after surgery (Mohseni et al., 2012). Non-invasive characterization of functional connectivity during STN-DBS for PD is of particular interest, as it allows to study the higher-order circuit dynamics and mechanism of action. Using functional MRI (fMRI), STN-DBS was found to desensitize the STN of its afferent, while simultaneously strengthening corticostriatal, thalamocortical, and direct pathways (Kahan et al., 2014).

Although the neurochemical correlates are not precisely established in humans, preclinical studies in a 6-hydroxydopamine-lesioned PD rat model found that STN-DBS for 1 h increased relative synaptic connectivity among corticostriatal glutamatergic terminals, thereby potentially contributing to the therapeutic benefit by partially rescuing striatal glutamatergic pathways (Walker et al., 2012). STN-DBS in hemi-parkinsonian rats showed duration-dependent metabolic effects for up to 1 week, such as counteracting lesion-induced glutamate/glutamine/GABA increases in the striatum, and normalizing GABA in the SNr, concomitant with a therapeutic effect on motor phenotype (Melon et al., 2015). Transcranial magnetic stimulation (TMS) is another useful non-invasive method to study long- and short-term plasticity during DBS, via paired-pulse stimulation and motor evoked potential (MEP) recording. Short-interval intracortical inhibition associated with motor improvement has been observed in STN and GPi DBS for PD, suggesting a restoration of the thalamocortical pathway during stimulation (Pierantozzi et al., 2002; Kim et al., 2015). Other network effects include enhancement of cortical reactivity, suggesting possible antidromic excitation of fibers connecting STN and motor cortex (Hanajima et al., 2004; Casula et al., 2017).

Structural and functional connectivity studies have begun to illuminate the importance of network effects beyond the target nucleus and basal ganglia circuit. A recent study investigated connectivity as a predictor of DBS outcome by using retrospective clinical data from PD patients who underwent STN-DBS at two centers. The authors used a publicly available connectome database acquired via diffusion tractography and resting-state functional connectivity to generate a connectivity profile for each patient (Horn et al., 2017). They found that certain connectivity profiles reliably predicted clinical outcome independent of electrode location, suggesting that network-wide phenomena might be as important as local molecular effects (Horn et al., 2017). Specifically, connectivity from the DBS target to the supplementary motor area and primary motor cortex was highly predictive for clinical outcome, corroborating evidence in TMS studies targeting these cortical regions (Shirotta et al., 2013; Brys et al., 2016; Horn et al., 2017). Further prospective studies are needed that examine the relationship between electrode location, anatomic and functional connectivity, and disease-relevant clinical outcomes.

### Epigenetic effects

DNA-methylation status in brain and blood samples from PD patients show high levels of congruence (Consales et al., 2018). Thus, a transcription fingerprint of leukocytes from PD patients was proposed, as these have been shown to respond to different brain pathologies such as stroke (Soreq et al., 2013). A distinct pattern of microRNAs between healthy subjects, PD patients under medical treatment, and PD patients with DBS was observed (Soreq et al., 2013). Changes in the methylation pattern were even observed after 1 h without stimulation, suggesting rapid miRNA responses (Soreq et al., 2013) related to inflammatory control and protection against DNA damage. The authors suggest that leukocyte methylation patterns could be used as biomarkers for effective DBS treatment of PD. The practical clinical implications might be individualized and adjustable DBS therapy based on peripheral epigenetic fingerprints, much in the same way that serial neurological assessment is used to titrate DBS contacts and voltage (Mohammadi & Meh dizadeh, 2016; Consales et al., 2018). Nevertheless, clinical application is likely years away, as further validation of epigenetic biomarkers is needed to determine the sensitivity, specificity, and clinical correlates of such peripheral measurements.

### Neurogenesis, synaptogenesis, and neuroprotection

Growing preclinical evidence suggests that, in addition to alleviating symptoms, STN-DBS has disease-modifying effects such as slowing nigral degeneration and neuroprotection. Rodent models have shown that chronic STN-DBS can lead to reduced loss of tyrosine-hydroxylase-positive (TH+) nigral neurons compared to sham stimulation (Temel et al., 2006; Spieles-Engemann et al., 2010; Fischer et al., 2017; Musacchio et al., 2017). Tyrosine hydroxylase is essential for metabolizing l-tyrosine to levodopa. In line with these observations, Wallace et al. tested STN-DBS on non-human primates with acute bilateral MPTP-mediated (methylphenyl-tetrahydropyridine) SNC TH+ lesions; the animals received either unilateral STN lesions or DBS stimulation for a week. The authors found 20–24% more TH+ DA cells in the SNC ipsilateral to stimulation or lesion, suggesting a protective effect of DBS (Wallace et al., 2007).

Recently, neurotrophic mechanisms of STN-DBS neuroprotection have also been demonstrated in preclinical models of PD (Spieles-Engemann et al., 2010; Fischer et al., 2017; Fischer & Sortwell, 2018). Specifically, in vivo studies showed that STN-DBS promotes expression of BDNF in the basal ganglia, which in turn binds to the tropomyosin-related kinase type 2 (trkB) receptor. As the chief BDNF and neurotropin-3 receptor in the basal ganglia, trkB has intermediate and long-term downstream effects including promoting dopaminergic neuron survival (Bayduk et al., 2011), regulating synaptic plasticity (Leal et al., 2014), and maintenance of the nigrostriatal pathway as a whole (Bayduk et al., 2011; Fischer et al., 2017). Increased BDNF was seen with high-frequency electrical stimulation of cultured dopaminergic neurons (Hartmann et al., 2001), as well as STN-DBS of PD rodent models (Fischer et al., 2017), with long-lasting...
neuroprotective benefits in the latter even after discontinuation of stimulation. Similarly, studies of STN-DBS in non-human primate models of PD have shown potential neuroprotective effects via inhibition of excitotoxic cell death (Tsukahara et al., 1995). Other proposed neuroprotective mechanisms include the following: promoting synaptogenesis via sustained trkB signaling (Guo et al., 2018) and facilitating striatal dopaminergic neurotransmission via maintenance of dendritic spine density (Rauskolb et al., 2010). Another theory, called electrotaxis, describes how the speed and directionality of precursor cell movement could be facilitated by the electric field generated from the DBS electrode (Jahanshahi et al., 2014).

Although post-mortem studies have found increased neural precursors in the subventricular zones of patients with chronic PD (Vedam-Mai et al., 2014), human trials to date have failed to show a direct disease-modifying effect of STN-DBS. Given that striatal dopaminergic dysfunction and neural loss are already well-established at the time of diagnosis, and that DBS is typically considered after 4 years of symptoms, it is perhaps unsurprising that surgical intervention has failed to produce any detectable disease modification in clinical trials. Moreover, PD patients enrolled in DBS trials are typically at that late stages of the disease, where the purpose of electrical stimulation is largely symptomatic. The design of future clinical trials should include more sensitive biomarkers and functional assessments of PD neuropathogenesis, and intervention at earlier stages where striatal neural salvage might still be possible (Fischer & Sortwell, 2018).

When applied to structures within the circuit of Papez, DBS has also been found to exert beneficial effects in neuropathological hallmarks, molecular expression, and behavior. Bilateral fornix DBS in the rat for 1-h induced expression of cFos, an immediate-early marker of neural activation, in the hippocampus. BDNF and VEGF were also significantly increased 2.5 h after stimulation, suggesting that neurotrophic and proliferating factors are associated with electrical stimulation (Gondard et al., 2015). Whether one would expect a constant increase in biochemical biomarkers before reaching a stable plateau or whether these markers show natural fluctuation under stimulation is so far not known. This might have future implications for cycled stimulation paradigms.

A similar influence on BDNF has been observed in a case study of chronic bilateral DBS of the lateral habenula for treatment-resistant depression: BDNF levels correlated in a reverse U-shape with levels of depression scores (Hoyer & Sartorius, 2012). Chronic fornix DBS was performed in a transgenic Alzheimer’s disease (AD) mouse model and showed rescue of memory impairment as well as amyloid-beta plaque clearance in the subiculum (Mann et al., 2017). The entorhinal cortex is another DBS target to rescue both early memory deficits and early plaque burden in a different AD mouse model, potentially mediated by direct projections from the entorhinal cortex to the hippocampus (Xia et al., 2017). Based on preliminary phase 1 studies in patients with improved cognition, reduction in cerebral atrophy, and cortical glucose metabolism positron emission tomography (PET) studies, an ongoing multi-national clinical trial of fornix DBS for AD is currently underway (Laxton et al., 2010; Sankar et al., 2014, 2015; Mirzadeh et al., 2016). Both fornix and entorhinal DBS have been shown to increase hippocampal neurogenesis in normal rodents (Stone et al., 2011) and disease models of Rett syndrome and AD.

Clinical effects of DBS

The cumulative effects of DBS therapy on the cellular, molecular, or electrical microlevel add up to clinically observable effects on the macrolevel for a variety of indications.

During the early years, DBS was applied to only the most severe cases of PD with a mean history of 11–13 years after diagnosis. As the therapy showed positive results, it was speculated if an earlier intervention might be beneficial. In a multicenter trial, 251 patients with only 7.5 years of history of PD and early motor complications were randomized to receive either the best medical therapy or the DBS. After 2 years, quality of life improved significantly in the DBS group, while it slightly decreased for patients with best medical treatment. Neurostimulation was also superior in regard to motor disability, activities of daily living, and levodopa-induced dyskinesias (Schuepbach et al., 2013). The levodopa dose was reduced by 39% in the DBS group while it was increased by 21% in the group with best medical treatment. Hypo- and hyperdopaminergic behavioral symptoms (like apathy or mania) were significantly reduced in the neurostimulation vs. the medical treatment group (Lhomée et al., 2018).

In the case of essential tremor, DBS has shown to lead to 60–75% long-term tremor control in more than 1,000 patients since 1998 (Dallapiazza et al., 2018). As these patients are for the most part compromised in daily activities (buttoning a shirt, holding a cup, using fork and knife, writing), it is no surprise that overall quality of life improved from 57.9 to 82% with DBS therapy in these patients (Dallapiazza et al., 2018).

Another standard indication for DBS is the treatment of dystonia. A meta-analysis of 523 patients with inherited or idiopathic dystonia showed a 65.2% reduction in dystonia motor rating scales after a mean 32.5 months of follow-up. Disability scores were reduced with DBS therapy by 58.6% (Mor o et al., 2017). As generalized dystonia especially affects pediatric patients, DBS represents an invasive but very effective therapy. A recent meta-analysis of 321 pediatric patients treated with DBS found that 86.3% showed a general and 66.1% a clinically significant improvement in dystonia scores (Elkaim et al., 2018).

In a multicenter study of therapy-refractory partial or secondarily generalized epilepsy, 110 patients received DBS in the anterior nucleus of the thalamus (ANT) and were randomized to active or inactive stimulation. After a 3-month blinded phase, the group with active DBS had a significantly higher reduction in seizure frequency (−40.4%) than the group without stimulation (−14.5%). After 2 years of active stimulation, the seizure frequency was reduced by 56% compared to baseline (Fisher et al., 2010). DBS therapy showed improvements of 47.7% in scores for 108 patients with obsessive–compulsive disorder and an overall responder rate (with a clinically meaningful reduction of 35% or more in the OCD score) of 58.2% (van Westen et al., 2015).

In summary, DBS positively affects the main symptoms of the corresponding disease and thereby reduces the amount of medication needed while increasing the quality of life for a large proportion of treated patients.

Current and future developments

A more complete understanding of the different mechanisms through which DBS exerts its action has opened up the field for new
Figure 7. Overview of selected targets for DBS.
Location of DBS targets for established and investigational indications: (A) DBS targets in coronal projection view (rostral to anterior commissure), (B) DBS targets in coronal projection view (anterior commissure to midcommissural point), (C) midcommissural point to posterior commissure), and (D) overview of thalamic DBS targets.
clinical indications, new stimulation targets, and new ways to deliver stimulation to the brain (Fig 7A–D). Various psychiatric circuitopathies have been explored with DBS therapy. Multiple targets such as the subgenual cingulate cortex (Brodemann Area 25, CG25; Holtzheimer et al., 2017), the NAcc (Bewernick et al., 2010; Fig 7A), the anterior limb of the internal capsule (ALIC; Bergfeld et al., 2016; Fig 7A), the medial forebrain bundle (Schlaepfer et al., 2013), and the lateral habenula (Sartorius et al., 2010) were investigated for treatment of MDD. DBS for treating OCD has been performed in the anterior limb of the internal capsule (Goodman et al., 2010), the bed nucleus of the stria terminalis (Nuttin et al., 2013), and the STN (Chabardès et al., 2013; Fig 7C) and the NAcc (Huff et al., 2010; Fig 7A). The NAcc has also been used to treat different forms of addiction (Müller et al., 2016). Tourette’s syndrome has been treated with DBS in the NAcc (Kuhn et al., 2007), globus pallidus internus (Akbarian-Tefaghi et al., 2017; Fig 7B), and the ventral anterior thalamus (Voa; Huys et al., 2016; Fig 7D). Eating disorders such as anorexia nervosa (CG25; Lipsman et al., 2013) or morbid obesity (with NAcc and lateral hypothalamic DBS; Whiting et al., 2013; Tronnier et al., 2018) have also been investigated as potential indications for DBS therapy. DBS of the fornix (Lozano et al., 2016; Fig 7A) and the nucleus basalis of Meynert (Hardenacke et al., 2013) is being explored as a treatment for Alzheimer’s. Combined stimulation of GPe and GPi (Fig 7B) is currently being explored for treating Huntington’s disease to reduce the amount of choreatic movements and cognitive decline (Wojtecki et al., 2016).

As the traditional indication for DBS therapy, movement disorders continually see new developments to improve effectiveness or efficiency. Adaptive closed-loop DBS systems can detect disease and state-specific biomarkers and adapt stimulation parameters delivered to the brain. This can include the power of beta oscillations in PD (Little et al., 2016) or accelerometer-detected amplitude in tremor (Herron et al., 2017). Other forms of stimulation rhythms that differ from current monophasic tonic stimulation are being explored, for example, with stimulation patterns similar to the synaptic noise generated by DBS (Leong et al., 2018).

Conclusion

Although the effects of DBS may often mimic a temporary lesion, the underlying mechanisms of action by far surpass simple inactivation of neurons. DBS has complex electrical effects on single neurons and neuronal networks, changes neurotransmitter concentrations and dynamics, and shapes the microenvironment including astrocytes, microglia, and endothelial cells. DBS also influences neuroplasticity and possibly induces neuroprotection and neurogenesis. With stereotactic implantation of electrodes with high precision and the possibility to modulate and titrate therapeutic effects, DBS is rapidly becoming a powerful tool to study brain function and physiology and an effective treatment for many different diseases.

Author contributions

MJ: manuscript draft, literature research, and figures; AF: manuscript draft and literature research; AML: correction; and KLK: conception of article, manuscript draft, and correction.

Conflict of interest

The authors declare that they have no conflict of interest.

Pending issues

(i) In vitro electrophysiology studies to refine electrical and neurochemical mechanisms
(ii) Effects of DBS on vasculature, blood–brain barrier, and glia (non-neural tissue)
(iii) Identifying optimal DBS targets for each indication
(iv) Refining parameters for DBS such as frequency, pulse width, and duration
(v) Prospective, randomized, and blinded trials on outcomes of novel indications for DBS
(vi) Closed-loop neuromodulation to individualize DBS.

References


Mechanisms of deep brain stimulation


STN-DBS share a common marker: reduced GABA content in the motor thalamus. Cell Death Dis 2: e154

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